2 DE as part of integrated proteomics: A full system's biology approach

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Summary
Despite a variety of new approaches, proteomics still relies heavily on two-dimensional electrophoresis as underlying separation technology. As functional and/or physical protein identification relies in all cases on secondary steps such as selective, multiplexed visualization, mass spectrometry and sequencing from cut spots, the need to view 2DE as a part of integrated proteomics became apparent. The challenge to apply a full systems biology approach in order to interpret the complexity of protein pattern and their interactions in the context of their metabolism or to detect and define biomarkers for diseases requires an unorthodox and new approach for knowledge management, information linking, sample tracking across instruments and laboratories and integration of interdisciplinary collaboration. The poster presents several “real life” examples applying a new commercially available “intelligent” desktop solution, which provides for integration of instruments across vendor barriers, existing applications and disparate data. In this “sentient” environment, the 2DE analysis module proofs not only orders of magnitude improvements in performance, but acts through active linking to relevant data and querying of public, subscription-based and client databases in the dynamically changing life sciences environment as a powerful systems biology approach to provide valuable, otherwise inaccessible scientific insights, sample tracking, experiment auditing and functional linking for any size laboratory. This new technology which utilizes vectorized n-dimensional data access methods and provides enhanced LIMS sample tracking functions, and its benefits for 2DE proteomics as a “system’s approach” are demonstrated and discussed.

Introduction
The complexity of proteomics’ functional data dependencies requires unorthodox approaches as integration of all available information is the key to effective knowledge management. While two-dimensional electrophoresis still remains a focal point in multi-parametric separation, the need for uniform and instant access to secondary analytical steps such as selective, multiplexed visualization, mass spectrometry (MS^n), protein sequencing, chemical structure and a broad variety of bioassays becomes mandatory. Additionally, interdisciplinary collaborations between life science research, clinical research, forensics and environmental science require an even more challenging effort.
This paper demonstrates the use of active data objects ("sentient", aware data) called "intelligent multidimensional objects" (IMO©) as smart agents which keep track of data history, processing steps, relationships between data and are able to communicate across formats and application boundaries. On examples of simultaneous queries from 2DE spots across global public, integrated client and subscription-service based database resources, the effective use of this technology to link instruments, applications and data in a flexible, content-relevant and dynamic fashion and treat them as a single, homogenous “virtual database” is demonstrated.

Materials & Methods

All shown results were obtained using a Dell Inspiron 8500 laptop, 512 MB RAM (Dell Corp., Ft. Lauderdale, FL) and an internet connection. Operating system was Windows XP Professional (Microsoft, Redmont, WA). Used software was Sentient Desktop™ 2.1 (IO Informatics Inc., Emeryville, CA.) as applications framework and Sentient 2DE module 2.1 (IO Informatics Inc., Emeryville, CA.) for 2D analysis. All screen captures were taken in wide (16:9) mode at 1680x1050 resolution, 32bit color, 96 dpi. Plug-in 3rd party tools shown in the examples were Weblab Viewer Lite 4.0 (Molecular Simulations, Inc.) and StarTree Viewer 1.1 (Inxight Software Inc.). Other tools used were publicly available parsers on the web like TagIdent (available for SwissProt/TrEMBL via ExPaSy, Swiss Institute of Bioinformatics).

Experiment 2DE image data were obtained from various experimental sources including public databases (SwissProt 2D, Swiss Institute of Bioinformatics). Populated data relationship views are courtesy of Dr. Frank Witzmann, IUPUI Indianapolis, IN. Spot detection, deconvolution and multi-parametric normalization for gel experiments with different gel sizes and visualization techniques were performed using Sentient 2DE module 2.1. Content definitions and environment settings were used from the default life science ontology without modifications.
Databases for real-time querying were used as pre-configured in the Sentient Desktop without any custom modifications. An overview about the currently included 150 public databases and their tree hierarchy is depicted in Fig. 3 shows the collapsed and Genomics/Proteomics expanded view.

Fig. 3: Life science database tree and expanded Genomics / Proteomics public databases

Results
A drag-&-drop query from a pl/molecular weight calibrated 2DE protein spot (sample: human, kidney) was used to generate a “universal” query across a subset of 90+ biological databases and to obtain “active” annotation. When clicked on the gel spot, these annotations allow for automatic launch of the linked corresponding information in its native application as data viewer as well as for hotlinks directly to web pages. In distinct difference to other approaches, this allows to also incorporate e.g. search scripts, sub-queries, etc. within the web link accounting for proper expansion when additional data becomes available. Fig. 4 demonstrates all procedural steps involved.
Fig. 4A:
Step 1: Drag the magnifier over the spot of interest on the calibrated gel image

Fig. 4B:
Step 2: The query is formulated. Select ranges for experimental parameters and choices about the scope of related content and sources

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Step 3: A list of relevant databases is displayed together with the range boundaries for experimental parameters

Step 4: From the pre-selected list, highlight those databases you want to output in your browser
Results from a simultaneous query to **92 databases** were obtained typically **within 90 seconds** and are displayed in the user’s browser of choice.

(Note: Response time varies based on quality of internet connection and availability of external database. Within the authors’ testing, the range of variation was between 48 sec. and 2 min. with automatic re-routing to mirrors on a DSL or T1 connection. The actual connection bandwidth matters only for result retrieval, not for sending).

Literature information was downloaded locally and saved as file to be available for future references. The file was then actively linked to the corresponding protein spot.

![Fig. 5: Creating active links to application and web pages](image)

After querying and linking, all screens shown in Fig. 6 are available in their respective viewers by a single mouse-click on the spot annotation.

![Fig. 6A: Literature information (launched in default application MS Word)](image)
Fig. 6B: InterPro search result

Fig. 6C: NiceView output of SwissProt
Fig. 6D: Sequence information in ProDom

Fig. 6E: 3D structure view in WebLab Viewer (automatically launched on click with protein-specific settings)
Content relationships can be explored via dendrograms based on individual experiment content definitions. Fig. 7 displays a small sample set of data using different methods and their CIM and single-link clustered dendrogram. Fig. 8 depicts a searchable tree-view of experiments in a laboratory environment using Inxight’s Star Tree Viewer as fully communicating, independent external application.

![Figure 7](image-url)

**Fig. 7:**
Data from different experimental methods in their relationship using a content-based dendrogram view for clustering.
Fig. 8A: Experiment linking: 2DE as part of integrated laboratory activities. The tree is searchable and interactively responding to view relationships between sets of experiments (Displayed data subset courtesy of Dr. Frank Witzmann, IUPUI Indianapolis, IN)
Fig. 8B: Experiment linking: Branching of individual MS spot data from 2DE as part of integrated laboratory activities. Same data tree as shown in Fig. 8A, interactively dragged (Courtesy: Dr. Frank Witzmann, IUPUI Indianapolis, IN)

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References


